



Impactos do estresse térmico na reprodução de fêmeas bovinas

Heat stress impact on bovine female reproduction

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Resumo

O estresse por calor (ET) ocorre quando a temperatura ambiente excede a zona de conforto térmico animal. Várias respostas corporais inespecíficas, capitaneadas pelos sistemas nervoso, neuroendócrino e imunológico são acionadas para manter a homeostase e resfriar o animal. O ET afeta o eixo hipotálamo-hipofisário-gonadal, comprometendo a liberação de gonadotrofinas, e promove o acúmulo de espécies reativas de oxigênio e proteínas anormais nas células ovarianas. Em resposta, as células ativam mecanismos antioxidantes e de reparação do DNA, que reduzem o metabolismo celular e aumentam as chances de sobrevivência; quando a reparação não é possível, acontece a apoptose. O ET impacta negativamente a produção de estradiol ovariano, o comportamento do estro, o desenvolvimento folicular, a competência dos oócitos e do embrião, as taxas de concepção, o estabelecimento e a manutenção da gravidez e até mesmo a eficiência reprodutiva da progênie. O combate ao ET inclui estratégias de combate ao aquecimento global progressivo e de manejo para resfriar os animais, e diminuir a produção de calor metabólico. O uso de biotecnologia reprodutiva e estratégias genéticas para gerar animais termotolerantes são também essenciais

Palavras-chave: Bovinos, Estresse térmico; Eficiência reprodutiva.

Abstract

Heat stress (HS), a harmful condition affecting animal production, reproduction, and welfare, occurs when an animal is exposed to temperatures that exceed its thermal comfort zone. Several nonspecific body responses involving neural, neuroendocrine, and immune systems are triggered to keep homeostasis in such conditions. These responses, primarily directed to cooling the body, also impact the hypothalamic-pituitary-gonadal axis, compromising the bovine female's release of gonadotropins. Heat stress also promotes reactive oxygen species accumulation in ovarian cells, impairing protein folding and refolding, triggering antioxidant and DNA protection mechanisms. These mechanisms, directed to reduce cell metabolism and increase survival chances, are not always sufficient to protect the cell and result in apoptosis. Heat stress's systemic and cellular consequences impact ovarian estradiol production, estrous behaviors, follicular development, oocytes and embryo competence, conception rates, pregnancy establishment and maintenance, and even the future reproductive efficiency of the progenies of cows exposed to HS during pregnancy. The combat of heat stress includes strategies to alleviate the effect of progressive global warming, management strategies to cool the animals, reduced metabolic heat, and methane production dietary approaches. The use of reproductive biotechs and genetic strategies to increase thermotolerant animals are also critical to overcoming the harmful effect of HS.

Keywords: Cattle; Heat stress; Reproductive efficiency.

Introduction

Heat stress (HS) occurs when an animal is exposed to temperatures above its thermal comfort zone (Ames, 1980). Additionally, ambient humidity, solar radiation, and wind exposure can also influence this response (Mader and Griffin, 2015).

HS has profound deleterious effects on animal production and reproduction (Roth, 2020) and based on predicted global warming, the harmful effects of HS on animals will be exacerbated (Masson-Delmotte et al., 2021). Therefore, a thorough understanding of the HS impacts on the reproductive

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physiology of the animals is required. This review summarizes major impacts of HS on female bovine reproduction.

Neuroendocrine responses to stress

Stressful event activates the hypothalamic-hypophyseal axis, promoting hypothalamic secretion of the corticotropin-releasing hormone (CRH) and vasopressin (VP), triggering the adenohipophyseal secretion of the adrenocorticotrophic hormone (ACTH) and β -endorphin. The ACTH acts on the adrenal cortex, stimulating synthesis of cortisol (Engler *et al.*, 1989) and progesterone (P_4) (Yoshida and Nakao, 2005; Maziero *et al.*, 2011). Through a negative feedback mechanism, cortisol reduces the release of CRH and ACTH, whereas ACTH and β -endorphin lessen the secretion of CRH. However, these regulatory mechanisms also impact the hypothalamic- hypophyseal-gonadal axis, reducing secretion of gonadotropin-releasing hormone (GnRH) by the hypothalamus and luteinizing hormone (LH) by the adenohipophysis, lessening, therefore, the synthesis of 17β -estradiol (17β -E₂) by ovarian follicles (Valsamakis *et al.*, 2019).

Heat stress systemic impacts

The maintenance of a high production level, accompanied by high feed intake and, consequently, a high rate of metabolic heat production, increases HS risk (Kadzere *et al.*, 2002). This phenomenon is particularly evident in *Bos taurus* dairy cows due to their higher production levels (Honig *et al.*, 2012). Lactating Holstein cows need a temperature humidity index (THI) < 72 to maintain thermoneutrality (Bohmanova *et al.*, 2007), while non-lactating Holstein cows remain in thermal comfort, even with THI ~ 75.4 (Ferrazza *et al.*, 2017).

Decreased dry matter intake (DMI) and increased respiratory rate are primary responses to HS (Ferrazza *et al.*, 2017). The increased respiratory rate is an excellent indicator of HS (Ferrazza *et al.*, 2017) and an essential thermoregulatory mechanism (McDowell *et al.*, 1976). However, its excessive increase is considered a signal of low HS tolerance (Pereira *et al.*, 2008). Additionally, HS increases water intake (Nardone *et al.*, 1997), standing time (Allen *et al.*, 2015), and sweating (Veissier *et al.*, 2018).

Heat stress also depresses the immune system. The increased plasma cortisol concentrations impair phagocytic cells and lymphocytes (Sgorlon *et al.*, 2012). Additionally, the increase in the Th1:Th2 ratio and plasma concentrations of transthyretin, tumor necrosis factor alpha (TNF α), and IL-6, well-known inflammatory biomarkers, characterize HS as a systemic inflammatory condition (Min *et al.*, 2016).

Cellular responses to heat stress

HS increases the intracellular concentration of reactive oxygen species (ROS) (Yu *et al.*, 2006; Gu *et al.*, 2015) that promotes accumulation of unfolded/misfolded proteins (Read and Schröder, 2021) and DNA damage (Srinivas *et al.*, 2019); ROS also activates the *NRF2* transcription factor for antioxidant genes, e.g., catalase, and superoxide dismutase 1, which are fundamental for ROS excess neutralization (Amin *et al.*, 2014).

To control the accumulation of unfolded/misfolded proteins, cells respond by increasing concentrations of chaperones, e.g., heat shock protein 60 (HSP60), HSP70, molecules with a primary function of supporting protein folding and refolding (Wang *et al.*, 2020), mitigating deleterious effects of ROS and preventing apoptosis (Bindu *et al.*, 2011). In response to DNA damage, the cell increases its P53 concentrations, which reduces metabolism and interrupts the cell cycle to focus on DNA repairs. However, if cell impairment is irreversible, P53 triggers apoptosis (Kruiswijk *et al.*, 2015).

All cellular alterations induced by HS were recently described in bovine granulosa cells (BGC) cultured *in vitro* (Alemu *et al.*, 2018; Wang *et al.*, 2020; Khan *et al.*, 2020). In these cells, HSP70 also reduces genes and protein expression of follicle stimulating hormone receptors and the aromatase, reducing 17β -E₂ (Li *et al.*, 2017).

Heat stress reproductive effects

Estrus expression and sexual behavior



HS severely impacts sexual behavior by decreasing physical activity, reducing mating and mating acceptance behaviors during estrus (Pennington *et al.*, 1985). Furthermore, increased plasma concentrations of ACTH and cortisol are also related to reduced estrus expression, as cortisol decreases cerebral sensitivity to 17β -E₂. Furthermore, synthesis of 17β -E₂ is also impaired due to reduced LH pulsatility (Hein and Allrich, 1992) and reduced activity of ovarian steroidogenic enzymes (Wolfenson *et al.*, 2000).

Reduced 17β -E₂ concentrations decrease estrus duration and intensity (Bolocan, 2009) and reduce vaginal mucosal hyperemia and mucus discharge (Schüller *et al.*, 2017). HS drastically reduces the estrus detection rate (De Rensis and Scaramuzzi, 2003), and consequently, pregnancy rate (Black *et al.*, 2018).

Follicular development and ovulation

Due to reduced LH pulsatility and direct effects of HS on theca and granulosa cells (Hein and Allrich, 1992), HS substantially impairs ovarian follicular development and oocyte and embryo competency (Wilson *et al.*, 1998ab; Wolfenson *et al.*, 2000). Compromised follicles produce less 17β -E₂ and inhibin, preventing control of plasma FSH and, consequently, delay follicle selection and deviation (Wolfenson *et al.*, 1995; Roth *et al.*, 2000). The delay reduces the probability of *B. taurus* cow having a follicle of ~12 mm, with full ovulatory capacity (Schüller *et al.*, 2017; Sartori *et al.*, 2001).

After a dairy cow is exposed to prolonged HS, conception rate (CR) will only return to normal in ~40 to 60 d (Morton *et al.*, 2007). Similarly, in cattle more tolerant to heat, e.g., Gir cows (*B. indicus*), even at 100 d after the HS episode, still had lower follicular steroidogenesis and oocyte incompetence (Torres-Júnior *et al.*, 2008).

Luteogenesis, luteal function and luteolysis

Cows kept permanently under HS initially have plasma P₄ concentrations similar to those in a thermoneutral (TN) environment, although there are lower P₄ concentrations between the 6 and 14 days after estrus (Howell *et al.*, 1994; Wolfenson *et al.*, 1988a; Alhussien *et al.*, 2018), probably due to HS effects on luteal development. However, when HS occurs after ovulation, plasma P₄ concentrations are similar in HS and TN cows (Wilson *et al.*, 1998b; Mogollón *et al.*, 2020), implying that post-ovulation HS does not affect luteal steroidogenesis or P₄ metabolism.

In vitro, endometrial explants from pregnant or non-pregnant Angus and Brangus cows collected 17 days post-estrus produced more PGF₂ α when exposed to HS (42 °C) than those kept under TN condition (39 °C) (Putney *et al.*, 1988, 1989). Additionally, oxytocin challenge increased PGF₂ α production of non-pregnant cow explants, independent of temperature, whereas in pregnant cows explants, an increase was only observed under HS (Putney *et al.*, 1988). In another *in vitro* study, HS increased PGF₂ α production of endometrial explants from non-pregnant Brangus cows on the 17th day of the cycle but did not compromise the capacity of the INT τ to halt this increase (Malayer *et al.*, 1990). In a recent study, cells from the endometrial stroma of non-pregnant cows had increased *in vitro* synthesis of PGF₂ α under HS conditions; in addition, TNF α potentiated this increase, but IFN τ suppressed it (Sakai *et al.*, 2021). However, in dairy cows, pregnant or not, on the 17th day post-estrus, 7.5 hours of HS did not alter plasma PGFM concentrations, indicating no increased PGF₂ α (Putney *et al.*, 1989).

Contradictory results were also reported *in vivo*. In two studies, HS increased luteal phase ~9 and ~2 days in cows and heifers, respectively (Wilson *et al.*, 1998a,b), implying delayed luteolysis. However, this was not confirmed in another study (Trout *et al.*, 1998), and in yet another, there were indications of premature luteolysis (Wolfenson *et al.*, 1988a). Lastly, in a study by our group, HS did not affect luteolytic responses in non-lactating Holstein cows given 5 or 12.5 mg of PGF₂ α , implying that HS did not alter PGF₂ α luteal sensitivity (Mogollón *et al.*, 2020).

Establishment and maintenance of pregnancy

Cows under HS have lower rates of fertilization (Sartori *et al.*, 2002) and conception (Schüller *et al.*, 2017; Roth, 2020). Furthermore, as 17β -E₂ are essential for expression of endometrial P₄ receptors (Martin *et al.*, 2008) and preparing the uterus for pregnancy (Binelli *et al.*, 2014), the low plasma 17β -E₂ observed during HS are likely insufficient to prepare the endometrium to support early embryo development



and implantation. Also, HS reduces luteogenesis, plasma P4, and pregnancy establishment and maintenance (Alhussien *et al.*, 2018). Besides being fundamental for maintenance of pregnancy, P4 also seems to be required for immune tolerance of the conceptus (Robinson and Klein, 2012). Importantly, in this review, the harmful impacts of HS upon oocytes and embryos will not be discussed. This subject was reviewed by Moura *et al.* (2021) and can be assessed in the Proceedings of this Congress.

Increased plasma concentrations of IL-2 and lower concentrations of IL-10 in cows under HS also seem to create unfavorable conditions for pregnancy maintenance (Alhussien *et al.*, 2018). In women, the anti-inflammatory cytokines produced by Th-2 (e.g., IL-10) create favorable conditions for implantation, whereas pro-inflammatory cytokines from Th-1 cells (e.g., IL-2) are harmful to paternal antigens (Nickerson *et al.*, 1994; Chatterjee *et al.*, 2014).

As pregnancy advances, cows are more affected by deleterious effects of HS. In late pregnancy, HS impairs precalving mammary development (Tao and Dahl, 2013) and milk production in subsequent lactation (Do Amaral *et al.*, 2009). Prepartum exposure to HS also delays the first breeding postpartum, reduces fertility, increases the calving to conception interval (Akbarinejad *et al.*, 2017), and raises the risk of passive immunity transfer failure (Tao *et al.*, 2012; Laporta *et al.*, 2017).

Control of heat stress

Decreasing deleterious effects of HS depends on addressing global heating, including reductions in CO₂ emissions by changing from fossil fuels to renewable energy sources (Tollefson, 2018). Also, as dairy cattle are responsible for 20% of global methane production, another greenhouse gas, an increased efficiency is also necessary, enabling sustained or increased production with fewer cows (Pryce and Bell, 2017).

The urgency and need for HS mitigating measures depend on the stage of the production cycle. In dairy cows, critical stages are the dry, transition, and lactation periods (Collier *et al.*, 1982; Wolfenson *et al.* 1988b). Holstein cows prefer shade areas (Schütz *et al.*, 2011), and even if such shade does not provide a THI < 72, it reduces respiratory rate and increases DMI (Abreu *et al.*, 2020).

As ambient temperature increases, animal cooling mechanisms change from evaporative to non-evaporative. Novel ambient cooling systems, integrating fans and water sprinklers, are highly efficient in supporting evaporative heat loss (Negrón-Pérez *et al.*, 2019). Increased evaporative heat loss, even when implemented for short intervals, before and after the artificial insemination, increased pregnancy rates in Holstein cows exposed to HS (Moghaddam *et al.*, 2009).

Feeding management can also be used to mitigate increased ambient temperature. Feeding animals in the coolest periods of the day can increase dry matter intake (Hicks *et al.*, 1989), whereas diets with less forage (Mader *et al.* 1999) and richer in fats and carbohydrates seem to reduce metabolic heat (West, 1999). Another nutritional approach to reduce impacts of HS and improve fertility is diets containing melatonin, chromium, immunomodulatory supplements, and antioxidants (Hall *et al.*, 2014; Negrón-Pérez *et al.*, 2019).

In addition, fixed-time artificial insemination protocols can also mitigate HS harmful effects on estrus behavior display, improve oocyte quality and increase service and pregnancy rates (De La Sota *et al.*, 1998). Furthermore, *in vitro* embryo production under thermoneutral conditions attenuates adverse effects of HS during initial development, enabling transfer of a morula or blastocyst that is more resistant to HS, increasing fertility (Hansen and Areéchiga, 1999).

Intense genetic selection for milk production reduced cattle HS tolerance due to a strong negative correlation (Pryce and Haile-Mariam, 2020). Therefore, global warming challenges the scientific community to pursue genetic improvement strategies that restore thermotolerance without impairing production (Scheper *et al.*, 2016).

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References

- Abreu AS et al.** Natural tree shade increases milk stability of lactating dairy cows during the summer in the subtropics. *Journal of Dairy Research*, v.87, p.444–447, 2020.
- Akbarinejad V, Gharagozlou F, Vojgani M.** Temporal effect of maternal heat stress during gestation on the fertility and anti-Müllerian hormone concentration of offspring in bovine. *Theriogenology*, v.99,



p.69–78, 2017.

Alemu TW et al. Oxidative and endoplasmic reticulum stress defense mechanisms of bovine granulosa cells exposed to heat stress. *Theriogenology*, v.110, p.130–141, 2018.

Alhussien MN et al. Effect of tropical thermal stress on peri-implantation immune responses in cows. *Theriogenology*, v.114, p.149–158, 2018.

Allen JD et al. Effect of core body temperature, time of day, and climate conditions on behavioral patterns of lactating dairy cows experiencing mild to moderate heat stress. *Journal of Dairy Science*, v.98, p.118–127, 2015.

Ames D. Thermal environment affects production efficiency of livestock. *BioScience*, v.30, p. 57–460, 1980.

Amin A et al. Bovine embryo survival under oxidative-stress conditions is associated with activity of the NRF2-mediated oxidative-stress-response pathway. *Molecular Reproduction and Development*, v.81, p. 497–513, 2014.

Bindu S et al. Translocation of heme oxygenase-1 to mitochondria is a novel cytoprotective mechanism against non-steroidal anti-inflammatory drug-induced mitochondrial oxidative stress, apoptosis, and gastric mucosal injury. *Journal of Biological Chemistry*, v.286, p.39387–39402, 2011.

Binelli M et al. The role of proestrus on fertility and postovulatory uterine function in the cow. *Animal Reproduction*, v.11, p.246–253, 2014.

Black R et al. Effects of heat stress on reproduction. *Journal of Dairy Science*, v. 6, p. 453–498, 2018.

Bohmanova J, Misztal I, Cole JB. Temperature-humidity indices as indicators of milk production losses due to heat stress. *Journal of Dairy Science*, v.90, p.1947–1956, 2007.

Bolocan E. Effects of heat stress on sexual behavior in heifers. *Zootehnie și Biotehnologii*, v.42, n.1, p.141–148, 2009.

Chatterjee P et al. Regulation of the anti-inflammatory cytokines interleukin-4 and interleukin-10 during pregnancy. *Frontiers in Immunology*, v.5, p.253, 2014.

Collier RJ et al. Effects of heat stress during pregnancy on maternal hormone concentrations, calf birth weight and postpartum milk yield of Holstein Cows. *Journal of Animal Science*, v.54, n.2, p.309–319, 1982.

De La Sota R et al. Efficacy of a timed insemination program in lactating dairy cows during summer heat stress. *Theriogenology*, v.49, p.761–770, 1998.

De Rensis F, Scaramuzzi RJ. Heat stress and seasonal effects on reproduction in the dairy cow - A review. *Theriogenology*, v.60, n.6, p.1139–1151, 2003.

Do Amaral BC et al. Heat-stress abatement during the dry period: Does cooling improve transition into lactation? *Journal of Dairy Science*, v.92, n.12, p.5988–5999, 2009.

Engler D et al. Studies of the secretion of corticotropin-releasing factor and arginine vasopressin into the hypophysial-portal circulation of the conscious sheep. *Neuroendocrinology*, v.49, p.367–381, 1989.

Ferrazza RA et al. Thermoregulatory responses of Holstein cows exposed to experimentally induced heat stress. *Journal of Thermal Biology*, v.66, p.68–80, 2017.

Gu ZT et al. Heat stress induced apoptosis is triggered by transcription-independent p53, Ca²⁺ dyshomeostasis and the subsequent Bax mitochondrial translocation. *Scientific Reports*, v. 5, 2015.

Hall LW et al. Evaluation of OmniGen-AF in lactating heat-stressed Holstein cows. 25th Annual Florida Ruminant Nutrition Symposium, v.15, p. 6–26, 2014.

Hansen PJ, Arechiga CF. Strategies for managing reproduction in the heat-stressed dairy cow. *Journal of Animal Science*, v.77, p.36–50, 1999.

Hein KG, Allrich RD. Influence of exogenous adrenocorticotrophic hormone on estrous behavior in cattle. *Journal of Animal Science*, v.70, p.243–247, 1992.

Hicks RB, Owens FN, Gill DR. Behavioral patterns of feedlot steers. Oklahoma State University Annual Science Research Report, v.1, p.94–105, 1989.

Honig H et al. Performance and welfare of high-yielding dairy cows subjected to 5 or 8 cooling sessions daily under hot and humid climate. *Journal of Dairy Science*, v.95, p.3736–3742, 2012.

Howell JL et al. Corpus luteum growth and function in lactating Holstein cows during spring and summer. *Journal of Dairy Science*, v.77, p.735–739, 1994.

Kadzere CT et al. Heat stress in lactating dairy cows: a review. *Livestock Production Science*, v.77, p.59–91, 2002.

Khan A et al. Cellular and molecular adaptation of bovine granulosa cells and oocytes under heat stress. *Animals*, v.10, p.1–14, 2020.

Kruiswijk F, Labuschagne CF, Vousden KH. P53 in survival, death and metabolic health: A lifeguard



- with a license to kill. *Nature Reviews Molecular Cell Biology*, v.16, p.393-405, 2015.
- Laporta J et al.** In utero exposure to heat stress during late gestation has prolonged effects on the activity patterns and growth of dairy calves. *Journal of Dairy Science*, v.100, p.2976-2984, 2017.
- Li H et al.** Lipopolysaccharide and heat stress impair the estradiol biosynthesis in granulosa cells via increase of HSP70 and inhibition of smad3 phosphorylation and nuclear translocation. *Cellular Signalling*, v.30, p.130-141, 2017.
- Mader T L et al.** Feedlot diet roughage level for Hereford cattle exposed to excessive heat load. *The Professional Animal Scientist*, v.15, p.53-62, 1999.
- Mader TL., Griffin D.** Management of cattle exposed to adverse environmental conditions. *Veterinary Clinics of North America - Food Animal Practice*, v.31, p.247-258, 2015.
- Malayer JR et al.** Regulation of heat shock-induced alterations in the release of prostaglandins by the uterine endometrium of cows. *Theriogenology*, v.34, p.219-230, 1990.
- Martin I et al.** Immunohistochemical detection of receptors for oestrogen and progesterone in endometrial glands and stroma during the oestrous cycle in Nelore (*Bos taurus indicus*) cows. *Reproduction in Domestic Animals*, v.43, p.415-421, 2008.
- Masson-Delmotte VP et al.** IPCC, 2021: Climate Change 2021: The Physical Science Basis. Contribution of Working Group I to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change. Cambridge University Press. In Press, v.1, 2021.
- Maziero RRD et al.** Ovarian function in cows submitted to acute stress during proestrus. *Livestock Science*, v.138, p.105-108, 2011.
- McDowell RE et al.** Effect of climate on performance of Holsteins in first lactation. *Journal of Dairy Science*, v.59, n. 5, p.965-971, 1976.
- Min L et al.** Long-term heat stress induces the inflammatory response in dairy cows revealed by plasma proteome analysis. *Biochemical and Biophysical Research Communications*, v.471, p.296-302, 2016.
- Moghaddam A et al.** Effects of short-term cooling on pregnancy rate of dairy heifers under summer heat stress. *Veterinary Research Communications*, v.33, p.567-575, 2009.
- Mogollón HDG et al.** Short communication: Heat stress does not affect induced luteolysis in Holstein cows. *Journal of Dairy Science*, v.103, p.5629-5633, 2020.
- Morton JM et al.** Effects of environmental heat on conception rates in lactating dairy cows: Critical Periods of Exposure. *Journal of Dairy Science*, v.90, p.2271-2278, 2007.
- Moura M.T. et al.** Heat stress impact on bovine female reproduction. *Proceedings of the International Symposium in Animal Reproduction*, 2021.
- Nardone A et al.** Composition of colostrum from dairy heifers exposed to high air temperatures during late pregnancy and the early postpartum period. *Journal of Dairy Science*, v.80, p.838-844, 1997.
- Negrón-Pérez VM et al.** Invited review: Management strategies capable of improving the reproductive performance of heat-stressed dairy cattle. *Journal of Dairy Science*, v.102, p.10695-10710, 2019.
- Nickerson et al.** Cytokines and the Th1/Th2 paradigm in transplantation. *Current Opinion in Immunology*, v.6, p.757-764, 1994.
- Pennington JA et al.** Sexual activity of Holstein cows: seasonal effects. *Journal of Dairy Science*, v.68, p.3023-3030, 1985.
- Pereira AMF et al.** Effect of thermal stress on physiological parameters, feed intake and plasma thyroid hormones concentration in Alentejana, Mertolenga, Frisian and Limousine cattle breeds. *International Journal of Biometeorology*, v.52, p.199-208, 2008.
- Pryce JE, Bell MJ.** The impact of genetic selection on greenhouse-gas emissions in Australian dairy cattle. *Animal Reproduction Science*, v.57, p.1451-1456, 2017.
- Pryce JE, Haile-Mariam M.** Symposium review: Genomic selection for reducing environmental impact and adapting to climate change. *Journal of Dairy Science*, v.103, p.5366-5375, 2020.
- Putney DJ et al.** Heat stress-induced alterations in the synthesis and secretion of proteins and prostaglandins by cultured bovine conceptuses and uterine endometrium. *Biology of Reproduction*, v.39, p.717-728, 1988.
- Putney DJ et al.** Modulation of uterine prostaglandin biosynthesis by pregnant and nonpregnant cows at day 17 post-estrus in response to in vivo and in vitro heat stress. *Animal Reproduction Science*, v.20, p.31-47, 1989.
- Read A, Schröder M.** The unfolded protein response: An overview. *Biology*, v.10, p.1-10, 2021.
- Robinson DP, Klein SL.** Pregnancy and pregnancy-associated hormones alter immune responses and disease pathogenesis. *Hormones and Behavior*, v.62, p.263-271, 2012.
- Roth ZVI.** Reproductive physiology and endocrinology responses of cows exposed to environmental heat



- stress - Experiences from the past and lessons for the present. *Theriogenology*, v.155, p.150–156, 2020.
- Sakai S et al.** Heat stress influences the attenuation of prostaglandin synthesis by interferon tau in bovine endometrial cells. *Theriogenology*, v.165, p.52–58, 2021.
- Sartori R et al.** Fertilization and early embryonic development in heifers and lactating cows in summer and lactating and dry cows in winter. *Journal of Dairy Science*, v.85, p.2803–2812, 2002.
- Sartori R et al.** Follicular deviation and acquisition of ovulatory capacity in bovine follicles. *Biology of Reproduction*, v.65, p.1403–1409, 2001.
- Scheper C et al.** Evaluation of breeding strategies for polledness in dairy cattle using a newly developed simulation framework for quantitative and Mendelian traits. *Genetics Selection Evolution*, v.48, p.1–11, 2016.
- Schüller et al.** Impact of heat stress on estrus expression and follicle size in estrus under field conditions in dairy cows. *Theriogenology*, v.102, p.48–53, 2017.
- Schütz KE et al.** Dairy cattle prefer shade over sprinklers: Effects on behavior and physiology. *Journal of Dairy Science*, v.94, p.273–283, 2011.
- Srinivas US et al.** ROS and the DNA damage response in cancer. *Redox Biology*, v.25, p.101084, 2019.
- Scorlon S et al.** Administration of botanicals with the diet regulates gene expression in peripheral blood cells of Sarda sheep during ACTH challenge. *Domestic Animal Endocrinology*. v.43, p.213–226, 2012.
- Tao S et al.** Effect of late-gestation maternal heat stress on growth and immune function of dairy calves. *Journal of Dairy Science*, v.95, p.7128–7136, 2012.
- Tao S, Dahl GE.** Invited review: Heat stress effects during late gestation on dry cows and their calves. *Journal of Dairy Science*, v.96, p.4079–4093, 2013.
- Tollefson J.** Clock ticking on climate action. *Nature*, v.562, n. 726, p.172–173, 2018.
- Torres-Júnior S et al.** Effect of maternal heat-stress on follicular growth and oocyte competence in *Bos indicus* cattle. *Theriogenology*, v.69, p.155–166, 2008.
- Trout JP et al.** Characteristics of the estrous cycle and antioxidant status of lactating Holstein cows exposed to heat stress. *Journal of Dairy Science*, v.81, p.1244–1250, 1998.
- Valsamakis et al.** Stress, female reproduction and pregnancy. *Psychoneuroendocrinology*, v.100, p.48–57, 2019.
- Veissier I et al.** Heat stress in cows at pasture and benefit of shade in a temperate climate region. *International Journal of Biometeorology*, v.62, p.585–595, 2018.
- Wang J et al.** Heat stress on calves and heifers: A review. *Journal of Animal Science and Biotechnology*, v.11, p.1–8, 2020.
- West, J.W.** Nutritional strategies for managing the heat-stressed dairy cow. *Journal of Animal Science*, v.77, p.13–35, 1999.
- Wilson SJ et al.** Effects of controlled heat stress on ovarian function of dairy cattle. 1. Heifers. *Journal of Dairy Science*, v.81, p.2132–2138, 1998a.
- Wilson SJ et al.** Effects of controlled heat stress on ovarian function of dairy cattle. 2. Lactating Cows. *Journal of Dairy Science*, v.81, p.2124–2131, 1998b.
- Wolfenson D et al.** Effect of heat stress on follicular development during the estrous cycle in lactating dairy cattle. *Biology of Reproduction*, v.52, p.1106–1113, 1995.
- Wolfenson D et al.** Hyperthermia and body energy store effects on estrous behavior, conception rate, and corpus luteum function in dairy cows. *Journal of Dairy Science*, v.71, p.3497–3504, 1988a.
- Wolfenson D et al.** Dry period heat stress relief effects on prepartum progesterone, calf birth weight, and milk production. *Journal of Dairy Science*, v. 71, p. 809–818, 1988b.
- Wolfenson D et al.** Impaired reproduction in heat-stressed cattle: basic and applied aspects. *Animal Reproduction Science*, v.60–61, p.535–547, 2000.
- Yoshida C, Nakao T.** Response of plasma cortisol and progesterone after ACTH challenge in ovariectomized lactating dairy cows. *Journal of Reproduction and Development*, v.51, p.99–107, 2005.
- Yu T et al.** Increased production of reactive oxygen species in hyperglycemic conditions requires dynamic change of mitochondrial morphology. *Proceedings of the National Acad Sciences*, v.103, p.2653–2658, 2006.
-